

NANOTHERAPY TO DELAY COGNITIVE IMPAIRMENT: USING COLLOIDAL NANOCARRIERS TO BLOCK AMYLOID- β -INDUCED DAMAGE IN BRAIN CELL MEMBRANES

DOI: 10.25177/JNMS.2.1.RV.532

Review

Received Date: 07th Jun 2019

Copy rights: © This is an Open access article distributed under the terms of International License.

Accepted Date: 27th Jun 2019Published Date: 04th July 2019

CORRESPONDENCE AUTHOR

Joseph S. D'Arrigo, Ph.D

Cavitation-Control Technology Inc., Farmington, CT 06032, USA

Email: cavcon@ntplx.net

CITATION

Joseph S. D'Arrigo, NANOTHERAPY TO DELAY COGNITIVE IMPAIRMENT: USING COLLOIDAL NANOCARRIERS TO BLOCK AMYLOID- β -INDUCED DAMAGE IN BRAIN CELL MEMBRANES (2019) SDRP Journal of Nanotechnology & Material Science 2(1)

ABSTRACT

Background: Numerous published studies indicate that microvascular endothelial dysfunction precedes cognitive decline in Alzheimer's disease, and that preservation of a healthy cerebrovascular endothelium can be an important therapeutic target.

Methods: By incorporating appropriate drug(s) into biomimetic (lipid cubic phase) nanocarriers, one obtains a multitasking combination therapeutic which targets certain cell-surface scavenger receptors, mainly class B type I (i.e., SR-BI), and crosses the blood-brain barrier. Documented similarities in lipid composition between high-density lipoproteins (HDL) and the biomimetic (nanoemulsion) nanocarrier particles can partially simulate or mimic the known heterogeneity (i.e., subpopulations or subspecies) of HDL particles. Moreover, the above-described type BI scavenger receptor (i.e., either SR-BI (rodent) and/or CLA-1 (human) orthologs) has been shown to be a multifunctional receptor able to bind a broad variety of ligands, including HDL and chylomicron remnants.

Results: Such colloidal-nanocarrier targeting allows for various Alzheimer's-related cell types to be simultaneously searched out, *in vivo*, for localized drug treatment. Using various lipids and their mixtures to form self-assembled non-lamellar nanostructures, it has continually been reported possible to successfully obtain stable colloidal dispersions of (liquid-crystalline) lipid cubic phases with well-defined particle size and morphology. In particular, within the range of self-assembled phases in model surfactant-like lipid systems, various investigators further emphasize that monoglyceride-based lyotropic liquid-crystalline phases are relatively unique owing to their rich polymorphism in water and potential application as drug nanocarriers.

Conclusion: This (colloidal-nanocarrier) *in vivo* targeting advantage may be particularly important for repurposing an FDA-approved drug, especially one which has shown the added ability to restore some cognitive functions in certain animal models of Alzheimer's disease (e.g., the anticancer drug bexarotene). Bexarotene (and several analogs or other candidate-repurposing drugs) up to now, by itself (i.e., without nanocarrier), displayed poor CNS penetration in human subjects.

Keywords: Alzheimer's disease, blood-brain barrier, cognitive aging, dementia, drug targeting, lipid cubic phases, nanoemulsion, scavenger receptors, SR-BI

BACKGROUND

A frequent co-morbidity of cerebrovascular pathology and Alzheimer's disease pathology has been observed over past decades. Accordingly, much evidence has been reported which indicates that microvascular endothelial dysfunction, due to cerebrovascular risk factors (e.g., atherosclerosis, obesity, diabetes, smoking, hypertension, aging), precedes cognitive decline in Alzheimer's disease and contributes to its pathogenesis. It is no surprise, therefore, that vascular brain lesions are very common in people over 70 years old, and recent reviews (e.g., [1,2]) provide much evidence that a large proportion of dementia cases may be attributable to cerebrovascular disease. As a result, vascular cognitive impairment and dementia (VCID) is the second leading cause of dementia behind Alzheimer's disease [4,10,11].

Endothelial Dysfunction as a Therapeutic Target for Cognitive Impairment

It has been reconfirmed in the current literature (see below) that receptor-mediated endocytosis/transcytosis via lipoprotein receptors, particularly scavenger receptors (including class B type I, i.e., SR-BI), remains a major route for drug delivery across the blood-brain barrier (BBB). Accordingly, endothelial-cell modulation and repair is feasible by pharmacological targeting [1,2,12-26] via SR-BI receptors (cf. [27-31]). Recently, Fung et al. [32] specifically reported that SR-BI mediates the uptake and transcytosis of HDL across brain microvascular endothelial cells (i.e., across the BBB). Since SR-BI has already been identified as a major receptor for HDL (with their major apolipoprotein (*apo*)A-I) as well as for the recently reviewed [1,2] "lipid-coated microbubble/nanoparticle-derived" (LCM/ND) nanoemulsion (see below), this multitasking lipid nanoemulsion can arguably serve as a targeted, apoA-I-based, (SR-BI mediated) therapeutic agent for common (late-onset) dementias [2,29,31,33-37] (cf. [38-43]).

This targeted-drug-delivery approach, using the proposed LCM/ND lipid nanoemulsion for treating the more common (late-onset) dementias, receives added impetus from continual findings of cerebrovascular

pathology [1,44-51] and an apparent *endothelium* dysfunction [2,28-36,52-61] in both Alzheimer's disease and its major risk factors [1,2,62-71]. Furthermore, this (intravenous) combination therapeutic would make it possible for various cell types, all potentially implicated in Alzheimer's disease (see [1,2] for reviews; cf. [72,73]), to be simultaneously sought out and better reached for localized drug treatment of brain tissue *in vivo* [73] (cf. [74]).

Consistent with the above considerations concerning endothelial dysfunction in Alzheimer's disease, ApoA-I has very recently been reported by Camacho et al. [75] to play a special role in cerebral amyloid angiopathy (CAA). CAA is observed in more than 90% of patients with Alzheimer's disease, although it additionally has an independent contribution to the cognitive deterioration associated with age. CAA can be neuropathologically classified as CAA type I or type II, where type I is characterized by amyloid- β deposition in cerebral capillaries (versus type II where cerebral capillaries are not involved). The human data (from autopsy brains of 20 post-mortem cases), reported by Camacho et al., revealed that the ApoA-I immunohistochemical staining was localized strongly toward capillary walls with CAA (i.e., especially toward CAA type I pathology) [75].

Also, as alluded to above (see Abstract), the previously documented similarities in lipid composition among HDL (as well as native low-density lipoproteins (LDL) and modified LDL) and LCM/ND nanoemulsion particles can partially simulate or mimic the known heterogeneity (i.e., subpopulations or subspecies) of HDL particles (see [73] for a review). Moreover, the above-described type BI scavenger receptor (i.e., either SR-BI (rodent) and/or CLA-1 (human) orthologs) has been shown to be a multi-functional receptor able to bind a broad variety of ligands, including HDL, LDL, oxidized LDL (OxLDL), very-low-density lipoproteins (VLDL), and chylomicron remnants. The presence of amphiphatic helices is a common feature of "exchangeable apolipoproteins", which are known to be the primary ligands (including notably apoA-I) for SR-BI [3]. In view of all the above considerations, SR-BI emerges

as the most plausible candidate (of all lipoprotein receptors) for major involvement in the enhanced endocytosis of LCM/ND nanoemulsion(s) for targeted drug delivery [73].

LCM/ND Nanoemulsion Type, Lipid Cubic Phases, and Biomimetic Nanocarriers

The self-assembling LCM/ND lipid nanoemulsion class comprises nonionic lipids exclusively (e.g., [76]) throughout its coated microbubble's and/or related nanoparticle's (i.e., related lipid polymorphs') supramolecular structures(s). This biobased lipid composition of LCM/ND nanoemulsions (i.e., comprising glycerides and cholesterol compounds) is similar to lipids contained in several types of plasma lipoproteins; accordingly, when these LCM/ND nanoemulsion particles are injected into the bloodstream, they likely acquire (i.e., bind) plasma apolipoprotein(s) – including notably apoA-I [73].

Importantly, monoglyceride is the largest single-lipid fraction (by wt. %) of the powdered solid lipid surfactants used to produce the (Filmix®) LCM/ND nanoemulsions [73]. As a group, monoglycerides exhibit different phase behaviors when they are exposed to water [77] (cf. [78-81]). In agreement with numerous other investigators, Kaasgaard and Drummond [82] also state that all these types of liquid-crystalline phases are frequently stable in excess water, which facilitates the preparation of nanoparticle dispersions and makes them suitable candidates for the encapsulation and controlled release of drugs (cf. [83-89]).

In particular, the self-assembly of varied and useful *dispersed cubic* phases (among other liquid-crystalline phases) depends heavily on the acyl chain length of the glycerides (primarily monoglycerides) placed in contact with water [73]. There is great interest to utilize these *dispersed cubic* phases for the administration of drugs, or for the formulation of new delivery systems [73,87]. The (lyotropic or solvent-induced) cubic liquid-crystalline phases may be classified into two distinct classes: *bicontinuous* cubic phases [90-94] and micellar or *discontinuous* (e.g., type *Fd3m*) cubic phases [89,90]. As reviewed by Garg et al. [77], monoglycerides spontaneously form

bicontinuous cubic phases upon the addition of water, are relatively insoluble (allowing the formation of colloidal dispersions of cubic phases), and are resistant to changes in temperature. Accordingly, lipid nanoparticles comprising interior liquid-crystalline structures of curved lipid membranes (i.e., dispersed cubic phases) have been used to solubilize, encapsulate, and deliver medications to disease areas within the body [77] (see also [73,95-105]).

In addition to the above-described category of various bicontinuous cubic phases, the other above-named category referred to as “micellar or discontinuous” cubic phases is also worthy of comment at this point. Of particular interest within this latter category is the well-studied micellar cubic structure of the type *Fd3m* (which is often denoted by the number Q²²⁷) (e.g., [89]). Luzzati and coworkers have reported that this *Fd3m* cubic phase evidently requires a heterogeneous mixture of polar lipids [91]: Using the lipid examples they cite (and the lipid classification system of Small [81]), this *Fd3m* phase apparently must include *both* at least one (sufficiently polar) *insoluble swelling amphiphilic* lipid (e.g., monoglyceride) and at least one (weakly polar) *insoluble nonswelling amphiphilic* lipid (e.g., diglyceride and/or cholesterol) (e.g., [3]; cf. [73]) in order to self-assemble properly in (excess) water. Hence, the *dispersed Fd3m* cubic phase can represent a lipid/water system which is particularly relevant to the earlier-described (Filmix®) LCM/ND lipid nanoemulsion formulation(s) on account of the fact that the patent claims describing the precise lipid composition of such nanoemulsion formulations (see especially Claim #1 in [76]) specifically include cholesterol and three categories of (saturated) glycerides, that is, tri-, di-, and monoglycerides (see [76]). In view of all the advantageous attributes of monoglycerides (recounted in the preceding paragraphs), and since (saturated) monoglyceride represents the largest single-lipid fraction of the LCM/ND lipid nanoemulsion type, the monoglyceride content probably plays a dominant role in supporting the evident long-term stability of the liquid-crystalline lipid nanoparticles in such nanoemulsions (see also [73] for a detailed review).

Besides certain glyceride-based liquid-crystalline systems displaying colloidal stability in excess water, the same important attribute has been documented for cholesterol and cholesterol esters – all of which are present in LCM/ND nanoemulsion formulations [73]. For example, cholesterol and its esters change the packing structure of lipids, and in high concentrations they are known to induce the formation of a liquid-crystal phase [106]. In addition, Kuntsche et al. [107,108] have prepared lipid nanoparticles in the (mesomorphic or) liquid-crystalline phase from cholesterol esters with saturated acyl chains. In accord with the above observations and considerations, the substantial concentrations of cholesterol esters and cholesterol in the LCM/ND nanoemulsion formulation likely further contribute to the known long-term stability of this nanoemulsion's (liquid-crystalline) lipid nanoparticles in excess water, thereby providing a persistent carrier matrix upon exposure to liquids such as blood plasma [73].

To conclude, self-assembled (colloidal mesophase) lipid nanoemulsions (e.g., [92-98]), particularly those predominantly containing dispersed cubic-phase lipid nanoparticles (e.g., [3,73,99-105]), continue to receive growing attention in pharmaceutical and/or biological fields. The main reason behind much of this attention is the fact that nonlamellar lipid nanostructures, such as cubic liquid-crystalline phases, have wide potential as delivery systems for numerous drugs, cosmetics, and food applications (cf. [3]). A recurring example of a largely monoglyceride-based drug-delivery agent category is the multitasking LCM/ND nanoemulsion formulation (cf. above). In this particular targeted-delivery approach, the self-assembled “lipid particle” structure itself (upon intravenous injection of the LCM/ND nanoemulsion) is apparently successfully utilized as the “active” targeting ligand – which is directed via (adsorption of) plasma lipoproteins toward the appropriate receptors on the target-cell surface. These dispersed liquid-crystalline lipid particles, of the LCM/ND nanoemulsion formulation, are colloidally stable nanocarriers which very likely represent liquid-crystalline inverse-topology nanotransporters (nanocarriers), i.e., dispersed lipid cubic phases (cf. [73]).

Amyloid- β Ion Channel Hypothesis of Alzheimer's Disease

As explained in many reviews (e.g., [109,110]) by different investigators, it has been recognized for over two decades that disturbance of the intracellular calcium homeostasis is central to the pathophysiology of several neurodegenerative disorders. As concerns Alzheimer's disease, it is believed by many researchers that enhanced calcium load may be brought about by extracellular accumulation of amyloid- β (A β) in the brain. Such studies have laid the foundation for the popular idea that amyloid- β peptides (39-42 amino acid molecules) are, in part, toxic to brain tissue because they form aberrant ion channels in cellular membranes and thereby disrupt Ca²⁺ homeostasis in brain tissue and increase intracellular Ca²⁺. More specifically, later studies indicated that soluble forms of A β facilitate influx through calcium-conducting ion channels in the plasma membrane, leading to excitotoxic neurodegeneration [109,110].

Historical support for the above amyloid- β ion channel hypothesis, or so-called “calcium hypothesis”, has also been observed at the clinical level [111]. Namely, there is little correlation between the amounts of fibrillar (insoluble) deposit at autopsy and the clinical severity of Alzheimer's disease. In contrast, a good correlation exists between early cognitive impairment and levels of soluble forms of A β in the brain [112]. (Aggregation of A β proceeds from formation of soluble (low molecular weight) spherical oligomers toward eventually assuming a final and stable conformation as insoluble fibrils from which amyloid- β plaques are constituted. Neurotoxicity is associated with soluble aggregates (i.e., oligomers) of A β rather than with the plaques themselves.) Accordingly, related experimental work has already shown that application of soluble A β oligomers (but not monomers or fibrils) to cultured neuroblastoma cells evoked large increases in cytosolic calcium that arise largely through Ca²⁺ influx across the plasma membrane [112].

As summarized by Di Scala et al. [111], the structure of amyloid pores has been extensively studied by ul-

trastructural methods. In particular, one group of investigators recently applied strategies (widely used to examine the structure of membrane proteins) to study the two major A β variants, namely, A β (1-40) and A β (1-42). Under the optimized detergent micelle conditions: 1) A β (1-40) aggregated into amyloid fibrils; 2) contrariwise, A β (1-42) assembled into oligomers that inserted into lipid bilayers as well-defined pores [113]. (These amyloid pores adopted characteristics of a β -barrel arrangement.) Because A β (1-42), relative to A β (1-40), has a more prominent role in Alzheimer's disease, the higher propensity of A β (1-42) to form β -barrel pore-forming oligomers is an indication of their importance in Alzheimer's disease [113]. Very recently, a different research group reported very similar findings [114]. As background for their study, these latter authors point out that: elevated A β (1-42) plasma levels have been correlated with the progression of late-onset forms of Alzheimer's disease; A β (1-42) is significantly more neurotoxic than A β (1-40) both *in vivo* and in neuronal cell culture; and memory impairment is believed to be driven by A β (1-42) disruption of long-term (hippocampal) potentiation. In accordance with these considerations, the authors' own detailed experimental data [114] indicated that A β (1-42) assemblies in oligomeric preparations form ion channels (in membranes excised from cells of neuronal origin). In contrast, A β (1-40) oligomers, fibrils, and monomers did not form channels. Moreover, ion-channel-conductance results suggested that A β (1-42) oligomers, but not monomers and fibrils, formed pore structures. The authors concluded that their findings demonstrate that only A β (1-42) contains unique structural features that facilitate membrane insertion and channel formation, now aligning ion channel formation with the neurotoxic effect of A β (1-42) compared to A β (1-40) in Alzheimer's disease [114].

Promise of Bexarotene (or analogs) to Inhibit Cognitive Decline in Humans

The preceding discussion of amyloid pore formation, in the cellular membranes of brain tissue, leads to another important consideration – the role of cholesterol. Namely, cholesterol is required for the assembly of amyloid pores formed by A β (1-42) [111].

Therefore, an amphipathic drug (such as bexarotene) which competes with cholesterol for binding to A β (1-42) would be capable of preventing oligomeric channel formation (at least *in vitro*). Such a strategy has already been contemplated for the treatment of Alzheimer's and other neurodegenerative diseases that involve cholesterol-dependent toxic oligomers [115]. However, when *oral* administration of bexarotene was employed subsequently in a Phase Ib (proof of mechanism) clinical trial [116], bexarotene displayed poor CNS penetration in normal human subjects. (Hence, the observed absence of an effect on A β metabolism was likely reflective of the low CNS-levels of bexarotene achieved [116](cf. [117])).

Nonetheless, at least two recently published reports (both in 2017) on bexarotene indicate a continuing interest in the ability of this FDA-approved (anticancer) drug to: 1) bind free A β peptide, as well as 2) bexarotene's previously reported positive effects in Alzheimer's-disease mouse models [118,119] (cf. [120,121]). Such past studies in animal models of Alzheimer's disease, concerning the beneficial effects of bexarotene, have also motivated a detailed analysis by Fantini et al. [122] to elucidate the mechanisms underlying the anti-Alzheimer properties of bexarotene in brain cells. These investigators demonstrated that bexarotene shares structural analogy with cholesterol: both bexarotene and cholesterol are amphipathic compounds, with a large apolar part consisting of a succession of hydrocarbon rings and a small polar headgroup (hydroxyl for cholesterol, carboxylate for bexarotene). Because it is the first drug that can both inhibit the binding of cholesterol to A β (1-42) and prevent calcium-permeable amyloid pore formation in the plasma membrane of brain cells, bexarotene might be considered as the prototype of a new class of anti-Alzheimer compounds [122]. (Note that because bexarotene shares structural analogy with cholesterol, and the above-described LCM/ND nanoemulsion contains substantial concentrations of cholesterol esters and cholesterol (see above), incorporation of the bexarotene molecule into the LCM/ND nanocarrier is expected to represent an uncomplicated, straightforward formulation procedure commercially.) Moreover, Casali et al. [123] have very recently reported

that treatment of an Alzheimer's-disease mouse model with (this FDA-approved anticancer drug) bexarotene resulted in enhanced cognition in the APP/PS1 mice which resembled earlier findings. Strikingly, the authors observed sustained cognitive improvements in the mice even when bexarotene treatment was discontinued for 2 weeks. Also, they observed a sustained reduction in microgliosis and plaque burden, following drug withdrawal, exclusively in the hippocampus. Casali et al. concluded that bexarotene selectively modifies aspects of neuroinflammation in a region-specific manner to reverse hippocampal-dependent cognitive deficits in Alzheimer's-disease (APP/PS1) mice [123].

Additional molecular aspects, concerning the membrane-related mechanisms for the known neuroprotective effect, of bexarotene action on brain tissue continue to be suggested and/or described in the recent literature (cf. [124,125]). In the most recently published study, Kamp et al. [126] show by NMR and CD spectroscopy that bexarotene directly interacts with the transmembrane domain of the amyloid precursor protein (APP) in a region where cholesterol binds. (Note that A β peptides are derived from APP, by the sequential action of β - and γ -secretases. γ -Secretase cleavage occurs in the transmembrane domain, of the C-terminal fragment left by β -secretase cleavage, and results in the release of A β peptides of various lengths [126]. The longer, neurotoxic, A β (1-42) peptide is highly aggregation prone and represents the major A β species deposited in the brain [126-129]. Cholesterol promotes A β (1-42) aggregation by enhancing its primary nucleation rate by up to 20-fold [129].) Kamp et al. argue that their data [126] suggest that bexarotene is a pleiotropic molecule that interferes with A β metabolism through multiple mechanisms. More specifically, earlier work by Di Scala et al. [115] provided evidence that bexarotene competed with cholesterol for binding to A β and prevented oligomeric channel formation. Di Scala et al. argue that their findings indicate that it is possible to prevent the generation of neurotoxic oligomers by targeting the cholesterol-binding domain of A β peptides [115]. Note that such blocking of amyloid- β -induced neurotoxic pore formation can be expected

to avoid exacerbation of blood-brain barrier breakdown, already occurring at lower levels in aged humans with cognitive decline [130], and thereby prevent reaching higher levels of BBB breakdown associated with cognitive impairment (and/or eventually dementia) in late-onset Alzheimer's disease [130-132].

CONCLUSION

By incorporating the appropriate drug(s) into biomimetic (lipid cubic phase) nanocarriers, one obtains a multitasking combination therapeutic which targets certain cell-surface scavenger receptors, mainly class B type I (SR-BI), and crosses the BBB. Such biomimetic-nanoemulsion targeting allows for various Alzheimer's-related cell types to be simultaneously searched out, *in vivo*, for localized drug treatment. The proposed multitasking nanocarrier therapeutic appears likely to display greater efficacy at different stages of Alzheimer's disease. This (colloidal-nanocarrier) *in vivo* targeting advantage may be particularly important for repurposing an FDA-approved drug, especially one (such as the anticancer drug bexarotene) which has shown the added ability to restore some cognitive functions in certain animal models of Alzheimer's disease.

ACKNOWLEDGEMENTS

This research did not receive any specific grant from funding agencies in the public, commercial, or non-profit sectors.

Conflicts of Interest

The authors declare no conflict of interest. J.S.D. is employed at Cav-Con Inc.

REFERENCES

- [1] D'Arrigo, J.S. Alzheimer's disease, brain injury, and CNS nanotherapy in humans: Sonoporation augmenting drug targeting. *Med. Sci.* 2017, 5, 29. PMCID:PMC5753658 [View Article](#)
- [2] D'Arrigo, J.S. Nanotherapy for Alzheimer's disease and vascular dementia: Targeting senile endothelium. *Adv. Colloid Interface Sci.* 2018, 251, 44-54. PMID:29274774 [View Article](#) [PubMed/NCBI](#)
- [3] D'Arrigo, J.S. Targeting early dementia: Using lipid cubic phase nanocarriers to cross the blood-brain barrier. *Biomimetics* 2018, 3, 4. PMID:31105226 [View Article](#) [PubMed/NCBI](#)

- [4] Dichgans, M.; Leys, D. Vascular cognitive impairment. *Circ. Res.* 2017, 120, 573-591. PMid:28154105 [View Article](#) [PubMed/NCBI](#)
- [5] Greenberg, S.M. Vascular disease and neurodegeneration: Advancing together. *Lancet Neurol.* 2017, 16, 333. 30086-8 [View Article](#)
- [6] Kalaria, R.N. Neuropathological diagnosis of vascular cognitive impairment and vascular dementia with implications for Alzheimer's disease. *Acta Neuropathol.* 2016, 131, 659-685. PMid:27062261 [View Article](#) [PubMed/NCBI](#)
- [7] Duncombe, J.; Kitamura, A.; Hase, Y.; Ihara, M.; Kalaria, R.N.; Horsburgh, K. Chronic cerebral hypoperfusion: A key mechanism leading to vascular cognitive impairment and dementia. Closing the translational gap between rodentmodels and human vascular cognitive impairment and dementia. *Clin. Sci.* 2017, 131, 2451-2468. PMid:28963120 [View Article](#) [PubMed/NCBI](#)
- [8] Perrotta, M.; Lembo, G.; Carnevale, D. Hypertension and dementia: Epidemiological and experimental evidence revealing a detrimental relationship. *Int. J. Mol. Sci.* 2016, 17, 347. PMid:27005613 [View Article](#) [PubMed/NCBI](#)
- [9] Sudduth, T.L.; Weekman, E.M.; Price, B.R.; Gooch, J.L.; Woolums, A.; Norris, C.M.; Wilcock, D.M. Time-course of glial changes in the hyperhomocysteinemia model of vascular cognitive impairment and dementia (VCID). *Neuroscience* 2017, 341, 42-51. PMid:27890830 [View Article](#) [PubMed/NCBI](#)
- [10] Bhat, N.R. Vasculoprotection as a convergent, multi-targeted mechanism of anti-AD therapeutics and interventions. *J. Alzheimers Dis.* 2015, 46, 581-591. PMid:26402511 [View Article](#) [PubMed/NCBI](#)
- [11] Alzheimer's Disease International. World Alzheimer Report 2016; Alzheimer's Disease International: London, UK, 2016. Available online: www.alz.co.uk/worldreport2016 (accessed on 20 February 2018).
- [12] Srimanee, A.; Regberg, J.; Hallbrink, M.; Vajragupta, O.; Langel, U. Role of scavenger receptors in peptide-based delivery of plasmid DNA across a blood-brain barrier model. *Int. J. Pharm.* 2016, 500, 128-135. PMid:26773601 [View Article](#) [PubMed/NCBI](#)
- [13] De Boer, A.G.; van der Sandt, I.C.J.; Gaillard, P.J. The role of drug transporters at the blood-brain barrier. *Annu. Rev. Pharmacol. Toxicol.* 2003, 43, 629-656. PMid:12415123 [View Article](#) [PubMed/NCBI](#)
- [14] Almer, G.; Mangge, H.; Zimmer, A.; Prassl, R. Lipoprotein-related and apolipoprotein-mediated delivery systems for drug targeting and imaging. *Curr. Med. Chem.* 2015, 22, 3631-3651. PMid:26180001 [View Article](#) [PubMed/NCBI](#)
- [15] Preston, J.E.; Abbott, J.; Begley, D.J. Transcytosis of macromolecules at the blood-brain barrier. *Adv. Pharmacol.* 2014, 71, 147-163. PMid:25307216 [View Article](#) [PubMed/NCBI](#)
- [16] Di Marco, L.Y.; Venneri, A.; Farkas, E.; Evans, P.C.; Marzo, A.; Frangi, A.F. Vascular dysfunction in the pathogenesis of Alzheimer's disease-A review of endothelium-mediated mechanisms and ensuing vicious circles. *Neurobiol. Dis.* 2015, 82, 593-606. PMid:26311408 [View Article](#) [PubMed/NCBI](#)
- [17] Salmina, A.B.; Inzhutova, A.I.; Malinovskaya, N.A.; Petrova, M.M. Endothelial dysfunction and repair in Alzheimer-type neurodegeneration: Neuronal and glial control. *J. Alzheimers Dis.* 2010, 22, 17-36. PMid:20847414 [View Article](#) [PubMed/NCBI](#)
- [18] Tong, X.K.; Hamel, E. Simvastatin restored vascular reactivity, endothelial function and reduced string vessel pathology in a mouse model of cerebrovascular disease. *J. Cereb. Blood Flow Metab.* 2015, 35, 512-520. PMid:25564230 [View Article](#) [PubMed/NCBI](#)
- [19] Carradori, D.; Gaudin, A.; Brambilla, D.; Andrieux, K. Application of nanomedicine to the CNS diseases. *Int. Rev. Neurobiol.* 2016, 130, 73-113. PMid:27678175 [View Article](#) [PubMed/NCBI](#)
- [20] Koster, K.P.; Thomas, R.; Morris, A.W.; Tai, L.M. Epidermal growth factor prevents oligomeric amyloid-induced angiogenesis deficits in vitro. *J. Cereb. Blood Flow Metab.* 2016, 36, 1865-1871. PMid:27634936 [View Article](#) [PubMed/NCBI](#)
- [21] Zenaro, E.; Piacentino, G.; Constantin, G. The blood-brain barrier in Alzheimer's disease. *Neurobiol. Dis.* 2016, 107, 41-56. PMid:27425887 [View Article](#) [PubMed/NCBI](#)
- [22] Qosa, H.; Mohamed, A.; Al Rihani, S.B.; Batarseha, Y.S.; Duong, Q.V.; Keller, J.N.; Kaddoumi, A. High-throughput screening for identification of blood-brain barrier integrity enhancers: A drug repurposing opportunity to rectify vascular amyloid toxicity. *J. Alzheimers Dis.* 2016, 53, 1499-1516. PMid:27392852 [View Article](#) [PubMed/NCBI](#)
- [23] Hostenbach, S.; D'haeseleer, M.; Kooijman, R.; De Keyser, J. The pathophysiological role of astrocytic endothelin-1. *Prog Neurobiol.* 2016, 144, 88-102. PMid:27132521 [View Article](#) [PubMed/NCBI](#)
- [24] Koizumi, K.; Wang, G.; Park, L. Endothelial dysfunction and amyloid-induced neurovascular alterations. *Cell. Mol. Neurobiol.* 2016, 36, 155-165. PMid:26328781 [View Article](#) [PubMed/NCBI](#)
- [25] Goldwaser, E.L.; Acharya, N.K.; Sarkar, A.; Godsey, G.; Nagele, R.G. Breakdown of the cerebrovasculature and blood-brain barrier: A mechanistic link between diabetes mellitus and Alzheimer's disease. *J. Alzheimers Dis.* 2016, 54, 445-456. PMid:27497477 [View Article](#) [PubMed/NCBI](#)
- [26] Bredesen, D.E. Reversal of cognitive decline: A novel therapeutic program. *Aging (Albany, NY)* 2014, 6, 707-717. PMid:25324467 [View Article](#) [PubMed/NCBI](#)
- [27] Mahringer, A.; Reichel, V.; Ott, M.; MacLean, C.; Reimold, I.; Hollnack-Pusch, E.; Fricker, G. Overcoming the blood brain barrier: The challenge of brain drug targeting. *J. Nanoneurosci.* 2012, 2, 5-19. [View Article](#)
- [28] Robert, J.; Button, E.B.; Stukas, S.; Boyce, G.K.; Gibbs, E.; Cowan, C.M.; Gilmour, M.; Cheng, W.H.; Soo, S.K.; Yuen, B.; et al. High-density lipoproteins suppress $\text{A}\beta$ -induced PBMC adhesion to human endothelial cells in bioengineered vessels and in monoculture. *Mol. Neurodegener.* 2017, 12,

60. PMid:28830501 [View Article](#) [PubMed/NCBI](#)
- [29] Vishnyakova, T.G.; Bocharov, A.V.; Baranova, I.N.; Chen, Z.; Remaley, A.T.; Csako, G.; Eggerman, T.L.; Patterson, A.P. Binding and internalization of lipopolysaccharide by CLA-1, a human orthologue of rodent scavenger receptor B1. *J. Biol. Chem.* 2003, 278, 22771-22780. PMid:12651854 [View Article](#) [PubMed/NCBI](#)
- [30] Darlington,D.; Li, S.; Hou, H.; Habib, A.; Tian, J.; Gao, Y.; Ehrhart, J.; Sanberg, P.R.; Sawmiller, D.; Giunta, B.; et al. Human umbilical cord blood-derived monocytes improve cognitive deficits and reduce amyloid-pathology in PSAPP mice. *Cell Transplant.* 2015, 24, 2237-2250. PMid:26230612 [View Article](#) [PubMed/NCBI](#)
- [31] Chang, E.H.; Rigotti, A.; Huerta, P. Age-related influence of the HDL receptor SR-BI on synaptic plasticity and cognition. *Neurobiol. Aging* 2009, 30, 407-419. PMid:17719144 [View Article](#) [PubMed/NCBI](#)
- [32] Fung, K.Y.; Wang, C.; Nyegaard, S.; Heit, B.; Fairn, G.D.; Lee,W.L. SR-BI mediated transcytosis of HDL in brain microvascular endothelial cells is independent of caveolin, clathrin, and PDZK1. *Front. Physiol.* 2017, 8, 841. PMid:29163190 [View Article](#) [PubMed/NCBI](#)
- [33] Robert, J.; Stukas, S.; Button, E.; Cheng, W.H.; Lee, M.; Fan, J.; Wilkinson, A.; Kulic, I.; Wright, S.D.; Wellington, C.L. Reconstituted high-density lipoproteins acutely reduce soluble brain A levels in symptomatic APP/PS1 mice. *Biochim. Biophys. Acta* 2016, 1862, 1027-1036. PMid:26454209 [View Article](#) [PubMed/NCBI](#)
- [34] Armstrong, S.M.; Sugiyama, M.G.; Fung, K.Y.Y.; Gao, Y.; Wang, C.; Levy, A.S.; Azizi, P.; Roufaiel,M.; Zhu, S.N.; Neculai, D.; et al. A novel assay uncovers an unexpected role for SR-BI in LDL transcytosis. *Cardiovasc. Res.* 2015, 108, 268-277. PMid:26334034 [View Article](#) [PubMed/NCBI](#)
- [35] Hottman, D.A.; Chernick, D.; Cheng, S.; Wang, Z.; Li, L. HDL and cognition in neurodegenerative disorders. *Neurobiol. Dis.* 2014, 72, 22-36. PMid:25131449 [View Article](#) [PubMed/NCBI](#)
- [36] Velagapudi, S.; Yalcinkaya, M.; Piemontese, A.; Meier, R.; Norrelykke, S.F.; Perisa, D.; Rzepiela, A.; Stebler, M.; Stoma, S.; Zanoni, P.; et al. VEGF-A regulates cellular localization of SR-BI as well as transendothelial transport of HDL but not LDL. *Arterioscler. Thromb. Vasc. Biol.* 2017, 37, 794-803. PMid:28360088 [View Article](#) [PubMed/NCBI](#)
- [37] Choi, H.J.; Seo, E.H.; Yi, D.; Sohn, B.K.; Choe, Y.M.; Byun, M.S.; Lee, J.M.; Woo, J.I.; Lee, D.Y. Amyloid-independent amnestic mild cognitive impairment and serum apolipoprotein A1 levels. *Am. J. Geriatr. Psychiatry* 2016, 24, 144-153. PMid:26238231 [View Article](#) [PubMed/NCBI](#)
- [38] Kitamura, Y.; Usami, R.; Ichihara, S.; Kida, H.; Satoh, M.; Tomimoto, H.; Murata, M.; Oikawa, S. Plasma protein profiling for potential biomarkers in the early diagnosis of Alzheimer's disease. *Neurol. Res.* 2017, 39, 231-238. PMid:28107809 [View Article](#) [PubMed/NCBI](#)
- [39] Lazarus, J.; Mather, K.A.; Armstrong, N.J.; Song, F.; Poljak, A.; Thalamuthu, A.; Lee, T.; Kochan, N.A.; Brodaty, H.; Wright, M.J.; et al. DNA methylation in the apolipoprotein-A1 gene is associated with episodic memory performance on healthy older individuals. *J. Alzheimers Dis.* 2015, 44, 175-182. PMid:25261444 [View Article](#) [PubMed/NCBI](#)
- [40] Ma, C.; Li, J.; Bao, Z.; Ruan, Q.; Yu, Z. Serum levels of apoA1 and apoA2 are associated with cognitive status in older men. *Biomed. Res. Int.* 2015, 2015, 481621. PMid:26682220 [View Article](#) [PubMed/NCBI](#)
- [41] Slot, R.E.; Van Harten, A.C.; Kester, M.I.; Jongbloed, W.; Bouwman, F.H.; Teunissen, C.E.; Scheltens, P.; Veerhuis, R.; van der Flier,W.M. Apolipoprotein A1 in cerebrospinal fluid and plasma and progression to Alzheimer's disease in non-demented elderly. *J. Alzheimers Dis.* 2017, 56, 687-697. PMid:28035918 [View Article](#) [PubMed/NCBI](#)
- [42] Yin, Z.G.; Li, L.; Cui, M.; Zhou, S.M.; Yu, M.M.; Zhou, H.D. Inverse relationship between apolipoprotein A-I and cerebral white matter lesions: A cross-sectional study in middle-aged and elderly subjects. *PLoS ONE* 2014, 9, e97113. PMid:24820970 [View Article](#) [PubMed/NCBI](#)
- [43] Weekman, E.M.; Sudduth, T.L.; Caverly, C.N.; Kopper, T.J.; Phillips, O.W.; Powell, D.K.; Wilcock, D.M. Reduced efficacy of anti-A immunotherapy in a mouse model of amyloid deposition and vascular cognitive impairment comorbidity. *J. Neurosci.* 2016, 36, 9896-9907. PMid:27656027 [View Article](#) [PubMed/NCBI](#)
- [44] Nelson, A.R.; Sweeney, M.D.; Sagare, A.P.; Zlokovic, B.V. Neurovascular dysfunction and neurodegeneration in dementia and Alzheimer's disease. *Biochim. Biophys. Acta* 2016, 1862, 887-900. PMid:26705676 [View Article](#) [PubMed/NCBI](#)
- [45] Kapasi, A.; Schneider, J.A. Vascular contributions to cognitive impairment, clinical Alzheimer's disease, and dementia in older persons. *Biochim. Biophys. Acta* 2016, 1862, 878-886. PMid:26769363 [View Article](#) [PubMed/NCBI](#)
- [46] McAleese, K.L.; Alafuzoff, I.; Charidimou, A.; De Reuck, J.; Grinberg, L.T.; Hainsworth, A.H.; Hortobagyi, T.; Ince, P.; Jellinger, K.; Gao, J.; et al. Post-mortem assessment in vascular dementia: Advances and aspirations. *BMC Med.* 2016, 14, 129. PMid:27600683 [View Article](#) [PubMed/NCBI](#)
- [47] Noh, Y.; Seo, S.W.; Jeon, S.; Lee, J.M.; Kim, J.S.; Lee, J.H.; Kim, J.H.; Kim, G.H.; Ye, B.S.; Cho, H.; et al. The role of cerebrovascular disease in amyloid deposition. *J. Alzheimers Dis.* 2016, 54, 1015-1026. PMid:27567803 [View Article](#) [PubMed/NCBI](#)
- [48] Hishikawa, N.; Fukui, Y.; Sato, K.; Kono, S.; Yamashita, T.; Ohta, T.; Deguchi, K.; Abe, K. Cognitive and affective functions in Alzheimer's disease patients with metabolic syndrome. *Eur. J. Neurol.* 2016, 23, 339-345. PMid:26493280 [View Article](#) [PubMed/NCBI](#)
- [49] Gutierrez, J.; Honig, L.; Elkind, M.S.; Mohr, J.P.; Goldman, J.; Dwork, A.J.; Morgello, S.; Marshall, R.S. Brain arterial aging and its relationship to Alz-

- heimer dementia. *Neurology* 2016, 86, 1507-1515. PMid:26984942 [View Article](#) [PubMed/NCBI](#)
- [50] Nagata, K.; Yamazaki, T.; Takano, D.; Maeda, T.; Fujimaki, Y.; Nakase, T.; Sato, Y. Cerebral circulation in aging. *Ageing Res. Rev.* 2016, 30, 49-60. PMid:27484894 [View Article](#) [PubMed/NCBI](#)
- [51] Calabrese, V.; Giordano, J.; Signorile, A.; Ontario, M.L.; Castorina, S.; de Pasquale, C.; Eckert, G.; Calabrese, E.J. Major pathogenic mechanisms in vascular dementia: Roles of cellular stress response and hormesis in neuroprotection. *J. Neurosci. Res.* 2016, 94, 1588-1603. PMid:27662637 [View Article](#) [PubMed/NCBI](#)
- [52] Toth, P.; Tarantini, S.; Csiszar, A.; Ungvari, Z.I. Functional vascular contributions to cognitive impairment and dementia: Mechanisms and consequences of cerebral autoregulatory dysfunction, endothelial impairment, and neurovascular uncoupling in aging. *Am. J. Physiol. Heart Circ. Physiol.* 2017, 312, H1-H20. PMid:27793855 [View Article](#) [PubMed/NCBI](#)
- [53] Devraj, K.; Poznanovic, S.; Spahn, C.; Schwall, G.; Harter, P.N.; Mittelbronn, M.; Antoniello, K.; Paganetti, P.; Muhs, A.; Heilemann, M.; et al. BACE-1 is expressed in the blood-brain barrier endothelium and is upregulated in a murine model of Alzheimer's disease. *J. Cereb. Blood Flow Metab.* 2016, 36, 1281-1294. PMid:26661166 [View Article](#) [PubMed/NCBI](#)
- [54] Chao, A.C.; Lee, T.C.; Juo, S.H.; Yang, D.I. Hyperglycemia increases the production of amyloid - peptide leading to decreased endothelial tight junction. *CNS Neurosci. Ther.* 2016, 22, 291-297. PMid:26842741 [View Article](#) [PubMed/NCBI](#)
- [55] Khalil, R.B.; Khoury, E.; Koussa, S. Linking multiple pathogenic pathways in Alzheimer's disease. *World J. Psychiatry* 2016, 6, 208-214. PMid:27354962 [View Article](#) [PubMed/NCBI](#)
- [56] Festoff, B.W.; Sajja, R.K.; van Dreden, P.; Cucullo, L. HGMB1 and thrombin mediate the blood-brain barrier dysfunction acting as biomarkers of neuroinflammation and progression to neurodegeneration in Alzheimer's disease. *J. Neuroinflamm.* 2016, 13, 194. PMid:27553758 [View Article](#) [PubMed/NCBI](#)
- [57] Gangoda, S.V.; Butlin, M.; Gupta, V.; Chung, R.; Avolio, A. Pulsatile stretch alters expression and processing of amyloid precursor protein in human cerebral endothelial cells. *J. Hypertens.* 2016, 34, e24. [View Article](#)
- [58] Roberts, A.M.; Jagadapillai, R.; Vaishnav, R.A.; Friedland, R.P.; Drinovac, R.; Lin, X.; Gozal, E. Increased pulmonary arteriolar tone associated with lung oxidative stress and nitric oxide in a mouse model of Alzheimer's disease. *Physiol. Rep.* 2016, 4, e12953. PMid:27604401 [View Article](#) [PubMed/NCBI](#)
- [59] Shang, S.; Yang, Y.M.; Zhang, H.; Tian, L.; Jiang, J.S.; Dong, Y.B.; Zhang, K.; Li, B.; Zhao, W.D.; Fang, W.G.; et al. Intracerebral GM-CSF contributes to transendothelial monocyte migration in APP/PS1 Alzheimer's disease mice. *J. Cereb. Blood Flow Metab.* 2016, 36, 1987-1991. PMid:27444968 [View Article](#)
- [60] Austin, S.A.; Katusic, Z.S. Loss of endothelial nitric oxide synthase promotes p25 generation and tau phosphorylation in amurine model of Alzheimer's disease. *Circ. Res.* 2016, 119, 1128-1134. PMid:27601478 [View Article](#) [PubMed/NCBI](#)
- [61] Katusic, Z.S.; Austin, S.A. Neurovascular protective function of endothelial nitric oxide. *Circ. J.* 2016, 80, 1499-1503. PMid:27238834 [View Article](#) [PubMed/NCBI](#)
- [62] Wang, L.; Du, Y.; Wang, K.; Xu, G.; Luo, S.; He, G. Chronic cerebral hypoperfusion induces memory deficits and facilitates A generation in C57BL/6J mice. *Exp. Neurol.* 2016, 283, 353-364. PMid:27421879 [View Article](#) [PubMed/NCBI](#)
- [63] Kyrtos, C.R.; Baras, J.S. Modeling the role of the glymphatic pathway and cerebral blood vessel properties in Alzheimer's disease pathogenesis. *PLoS ONE* 2015, 10, e0139574. PMid:26448331 [View Article](#) [PubMed/NCBI](#)
- [64] Kalaria, R.N.; Akinyemi, R.; Ihara, M. Stroke injury, cognitive impairment and vascular dementia. *Biochim. Biophys. Acta* 2016, 1862, 915-925. PMid:26806700 [View Article](#) [PubMed/NCBI](#)
- [65] Khan, A.; Kalaria, R.N.; Corbett, A.; Ballard, C. Update on vascular dementia. *J. Geriatr. Psychiatry Neurol.* 2016, 29, 281-301. PMid:27502303 [View Article](#) [PubMed/NCBI](#)
- [66] Austin, S.A.; Santhanam, A.V.; d'Uscio, L.V.; Katusic, Z.S. Regional heterogeneity of cerebral microvessels and brain susceptibility to oxidative stress. *PLoS ONE* 2015, 10, e0144062. PMid:26629821 [View Article](#) [PubMed/NCBI](#)
- [67] Toda, N.; Okamura, T. Cigarette smoking impairs nitric oxide-mediated cerebral blood flow increase: Implications for Alzheimer's disease. *J. Pharmacol. Sci.* 2016, 131, 223-232. PMid:27530818 [View Article](#) [PubMed/NCBI](#)
- [68] Uiterwijk, R.; Huijts, M.; Staals, J.; Rouhl, R.P.; De Leeuw, P.W.; Kroon, A.A.; van Oostenbrugge, R.J. Endothelial activation is associated with cognitive performance in patients with hypertension. *Am. J. Hypertens.* 2016, 29, 464-469. PMid:26271106 [View Article](#) [PubMed/NCBI](#)
- [69] Kamat, P.K.; Kyles, P.; Kalani, A.; Tyagi, N. Hydrogen sulfide ameliorates homocysteine-induced Alzheimer's disease-like pathology, blood-brain barrier disruption, and synaptic disorder. *Mol. Neurobiol.* 2016, 53, 2451-2467. PMid:26019015 [View Article](#) [PubMed/NCBI](#)
- [70] Iadecola, C. Untangling neurons with endothelial nitric oxide. *Circ. Res.* 2016, 119, 1052-1054. PMid:27789581 [View Article](#) [PubMed/NCBI](#)
- [71] Wang, Y.J. Lessons from immunotherapy for Alzheimer's disease. *Nat. Rev. Neurol.* 2014, 10, 188-189. PMid:24638135 [View Article](#) [PubMed/NCBI](#)
- [72] Krstic, D.; Knuesel, I. Deciphering the mechanism underlying late-onset Alzheimer's disease. *Nat. Rev. Neurol.* 2013, 9, 25-34. PMid:23183882 [View Article](#) [PubMed/NCBI](#)
- [73] D'Arrigo, J. Stable Nanoemulsions: Self-Assembly in Nature and Nanomedicine; Elsevier: Amsterdam,

- The Netherlands, 2011; 415p, ISBN 978-0-444-53798-0.
- [74] Barbarese, E.; Ho, S.Y.; D'Arrigo, J.S.; Simon, R.H. Internalization of microbubbles by tumor cells in vivo and in vitro. *J. Neurooncol.* 1995, 26, 25-34. PMid:8583242 [View Article](#) [PubMed/NCBI](#)
- [75] Camacho, J.; Moline, T.; Bonaterra-Pastrana, A.; Ramon y Cajal, S.; Martinez-Saez, E.; Hernandez-Guillamon, M. Brain apoA-I, apoJ and apoE immunodetection in cerebral amyloid angiopathy. *Front. Neurol.* 2019, 10, 187. PMid:30918495 [View Article](#) [PubMed/NCBI](#)
- [76] D'Arrigo, J. Surfactant Mixtures, Stable Gas-in-Liquid Emulsions, and Methods for the Production of such Emulsions from Said Mixtures. U.S. Patent No. 4,684,479A , 4 August 1987.
- [77] Garg, G.; Saraf, Sh.; Saraf, Sw. Cubosomes: An overview. *Biol. Pharm. Bull.* 2007, 30, 350-353. PMid:17268078 [View Article](#) [PubMed/NCBI](#)
- [78] Tanford, C. *The Hydrophobic Effect: Formation of Micelles and Biological Membranes*; Wiley: New York, NY, USA, 1973; 200p.
- [79] Boyd, B.J.; Whittaker, D.V.; Khoo, S.M.; Davey, G. Lyotropic liquid crystalline phases formed from glycerate surfactants as sustained release drug delivery systems. *Int. J. Pharm.* 2006, 309, 218-226. PMid:16413980 [View Article](#) [PubMed/NCBI](#)
- [80] Pouton, C.W. Properties and uses of common formulation lipids, surfactants and cosurfactants. In *Proceedings of the AAPSWorshop, Effective Utilization of Lipid-Based Systems to Enhance the Delivery of Poorly Soluble Drugs: Physicochemical, Biopharmaceutical and Product Development Considerations*, Bethesda, MD, USA, 5-6 March 2007; Constantinides, P.P., Porter, C.J.H., Eds.; AAPS: Arlington, VA, USA, 2007.
- [81] Small, D.M. The behavior of biological lipids. *Pure Appl. Chem.* 1981, 53, 2095-2103. [View Article](#)
- [82] Kaasgaard, T.; Drummond, C.J. Ordered 2-D and 3-D nano-structured amphiphile self-assembly materials stable in excess solvent. *Phys. Chem. Chem. Phys.* 2006, 8, 4957-4975. PMid:17091149 [View Article](#) [PubMed/NCBI](#)
- [83] Shearman, G.C.; Khoo, B.J.; Motherwell, M.L.; Brakke, K.A.; Ces, O.; Conn, C.E.; Seddon, J.M.; Templer, R.H. Calculations of and evidence for chain packing stress in inverse lyotropic bicontinuous cubic phases. *Langmuir* 2007, 23, 7276-7285. PMid:17503862 [View Article](#) [PubMed/NCBI](#)
- [84] Rizwan, S.B.; Dong, Y.D.; Boyd, B.J.; Rades, T.; Hook, S. Characterization of bicontinuous cubic liquid crystalline systems of phytantriol and water using cryo field emission scanning electron microscopy (cryo FESEM). *Micron* 2007, 38, 478-485. PMid:17011783 [View Article](#) [PubMed/NCBI](#)
- [85] Yaghmur, A.; de Campo, L.; Sagalowicz, L.; Leser, M.E.; Glatter, O. Emulsified microemulsions and oil-containing liquid crystalline phases. *Langmuir* 2005, 21, 569-577. PMid:15641825 [View Article](#) [PubMed/NCBI](#)
- [86] Yaghmur, A.; de Campo, L.; Sagalowicz, L.; Leser, M.E.; Glatter, O. Control of the internal structure of MLO-based isosomes by the addition of diglycerol monooleate and soybean phosphatidylcholine. *Langmuir* 2006, 22, 9919-9927. PMid:17106981 [View Article](#) [PubMed/NCBI](#)
- [87] Gustafsson, J.; Ljusberg-Wahren, H.; Almgren, M.; Larsson, K. Submicron particles of reversed lipid phases in water stabilized by a nonionic amphiphilic polymer. *Langmuir* 1997, 13, 6964-6971. + [View Article](#)
- [88] De Campo, L.; Yaghmur, A.; Sagalowicz, L.; Leser, M.E.; Watzke, H.; Glatter, O. Reversible phase transitions in emulsified nanostructured lipid systems. *Langmuir* 2004, 20, 5254-5261. PMid:15986660 [View Article](#) [PubMed/NCBI](#)
- [89] Yaghmur, A.; de Campo, L.; Salentinig, S.; Sagalowicz, L.; Leser, M.E.; Glatter, O. Oil-loaded monolinolein-based particles with confined inverse discontinuous cubic structure (Fd3m). *Langmuir* 2006, 22, 517-521. PMid:16401095 [View Article](#) [PubMed/NCBI](#)
- [90] Larsson, K. Aqueous dispersions of cubic lipid-water phases. *Curr. Opin. Colloid Interface Sci.* 2000, 5, 64-69. 00040-6 [View Article](#)
- [91] Luzzati, V. Biological significance of lipid polymorphism: The cubic phases. *Curr. Opin. Struct. Biol.* 1997, 7, 661-668. 80075-9 [View Article](#)
- [92] Sagar, G.H.; Arunagirinathan, M.A.; Bellare, J.R. Self-assembled surfactant nano-structures important in drug-delivery: A review. *Indian J. Exp. Biol.* 2007, 45, 133-159.
- [93] Anton, N.; Benoit, J.P.; Saulnier, P. Design and production of nanoparticles formulated from nano-emulsion templates: A review. *J. Control. Release* 2008, 128, 185-199. PMid:18374443 [View Article](#) [PubMed/NCBI](#)
- [94] Bansal, T.; Mustafa, G.; Khan, Z.I.; Ahmad, F.J.; Khar, R.K.; Talegaonkar, S. Solid self-nanoemulsifying delivery systems as a platform technology for formulation of poorly soluble drugs. *Crit. Rev. Ther. Drug Carrier Syst.* 2008, 25, 63-116. PMid:18540836 [View Article](#) [PubMed/NCBI](#)
- [95] Sadurni, N.; Solans, C.; Azemar, N.; Garcia-Celma, M.J. Studies on the formation of O/W nano-emulsions, by low-energy emulsification methods, suitable for pharmaceutical applications. *Eur. J. Pharm. Sci.* 2005, 26, 438-445. PMid:16153811 [View Article](#) [PubMed/NCBI](#)
- [96] Tresset, G. The multiple faces of self-assembled lipidic systems. *PMC Biophys.* 2009, 2, 3. PMid:19374753 [View Article](#) [PubMed/NCBI](#)
- [97] Hato, M.; Yamashita, J.; Shiono, M. Aqueous phase behavior of lipids with isoprenoid type hydrophobic chains. *J. Phys. Chem. B* 2009, 113, 10196-10209. PMid:19572621 [View Article](#) [PubMed/NCBI](#)
- [98] Barauskas, J.; Cervin, C.; Tiberg, F.; Johnsson, M. Structure of lyotropic self-assembled lipid nonlamellar liquid crystals and their nanoparticles in mixtures of phosphatidyl choline and -tocopherol (vitamin E). *Phys. Chem. Chem. Phys.* 2008, 10, 6483-6485. PMid:18979032 [View Article](#) [PubMed/NCBI](#)
- [99] Efrat, R.; Aserin, A.; Garti, N. On structural transitions in a discontinuous micellar cubic phase loaded

- with sodium diclofenac. *J. Colloid Interface Sci.* 2008, 321, 166-176. PMid:18279886 [View Article](#) [PubMed/NCBI](#)
- [100] Yaghmur, A.; Laggner, P.; Almgren, M.; Rappolt, M. Self-assembly in monoelaidin aqueous dispersions: Direct vesicles to cubosomes transition. *PLoS ONE* 2008, 3, e3747. PMid:19015726 [View Article](#) [PubMed/NCBI](#)
- [101] Yaghmur, A.; Glatter, O. Characterization and potential applications of nanostructured aqueous dispersions. *Adv. Colloid Interface Sci.* 2009, 147-148, 333-342. PMid:18804754 [View Article](#) [PubMed/NCBI](#)
- [102] Vandoolaeghe, P.; Rennie, A.R.; Campbell, R.A.; Nylander, T. Neutron reflectivity studies of the interaction of cubic phase nanoparticles with phospholipid bilayers of different coverage. *Langmuir* 2009, 25, 4009-4020. PMid:19714826 [View Article](#) [PubMed/NCBI](#)
- [103] Vandoolaeghe, P.; Barauskas, J.; Johnsson, M.; Tiberg, F.; Nylander, T. Interaction between lamellar (vesicles) and nonlamellar lipid liquid-crystalline nanoparticles as studied by time-resolved small-angle X-ray diffraction. *Langmuir* 2009, 25, 3999-4008. PMid:19714888 [View Article](#) [PubMed/NCBI](#)
- [104] Yaghmur, A.; Kriechbaum, M.; Amenitsch, H.; Steinhart, M.; Laggner, P.; Rappolt, M. Effects of pressure and temperature on the self-assembled fully hydrated nanostructures of monoolein-oil systems. *Langmuir* 2010, 26, 1177-1185. PMid:19681634 [View Article](#) [PubMed/NCBI](#)
- [105] Fong, W.K.; Hanley, T.; Boyd, B.J. Stimuli responsive liquid crystals provide "on-demand" drug delivery in vitro and in vivo. *J. Control. Release* 2009, 135, 218-226. PMid:19331865 [View Article](#) [PubMed/NCBI](#)
- [106] Amselem, S.; Friedman, D. Solid Fat Nanoemulsions. U.S. Patent No. 5,662,932A , 2 September 1997.
- [107] Kuntsche, J.; Koch, M.H.J.; Drechsler, M.; Bunjes, H. Crystallization behavior of supercooled smectic cholesteryl myristate nanoparticles containing phospholipids as stabilizers. *Colloids Surf. B Biointerfaces* 2005, 44, 25-35. PMid:15990283 [View Article](#) [PubMed/NCBI](#)
- [108] Kuntsche, J.; Westesen, K.; Drechsler, M.; Koch, M.H.J.; Bunjes, H. Supercooled smectic nanoparticles: A potential novel carrier system for poorly water soluble drugs. *Pharm. Res.* 2004, 21, 1834-1843. PMid:15553230 [View Article](#) [PubMed/NCBI](#)
- [109] Nimmrich, V.; Eckert, A. Calcium channel blockers and dementia. *Brit. J. Pharmacol.* 2013, 169, 1203-1210. PMid:23638877 [View Article](#) [PubMed/NCBI](#)
- [110] Shirwany, N.A.; Payette, D.; Xie, J.; Guo, Q. The amyloid beta ion channel hypothesis of Alzheimer's disease. *Neuropsychiatr. Dis. Treat.* 2007, 3, 597-612.
- [111] Di Scala, C.; Yahi, N.; Boutemeur, S.; Flores, A.; Rodriguez, L; Chahinian, H.; Fantini, J. Common molecular mechanism of amyloid pore formation by Alzheimer's β -amyloid peptide and α -synuclein. *Sci. Rep.* 2016, 6, 28781. PMid:27352802 [View Article](#) [PubMed/NCBI](#)
- [112] Demuro, A.; Smith, M.; Parker, I. Single-channel Ca²⁺ imaging implicates A β 1-42 amyloid pores in Alzheimer's disease pathology. *J. Cell Biol.* 2011, 195, 515-524. PMid:22024165 [View Article](#) [PubMed/NCBI](#)
- [113] Serra-Batiste, M.; Ninot-Pedrosa, M.; Bayoumi, M.; Gairi, M.; Maglia, G.; Carulla, N. A β 42 assembles into specific β -barrel pore-forming oligomers in membrane-mimicking environments. *Proc. Natl. Acad. Sci. USA* 2016, 113, 10866-10871. PMid:27621459 [View Article](#) [PubMed/NCBI](#)
- [114] Bode, D.C.; Baker, M.D.; Viles, J.H. Ion channel formation by amyloid- β 42 oligomers but not amyloid- β 40 in cellular membranes. *J. Biol. Chem.* 2017, 292, 1404-1413. PMid:27927987 [View Article](#) [PubMed/NCBI](#)
- [115] Di Scala, C.; Chahinian, H.; Yahi, N.; Garmy, N.; Fantini, J. Interaction of Alzheimer's β -amyloid peptides with cholesterol: Mechanistic insights into amyloid pore formation. *Biochemistry* 2014, 53, 4489-4502. PMid:25000142 [View Article](#) [PubMed/NCBI](#)
- [116] Ghosal, K.; Haag, M.; Verghese, P.B.; West, T.; Veenstra, T.; Braunstein, J.B.; Bateman, R.J.; Holtzman, D.M.; Landreth, G.E. Arandomized controlled study to evaluate the effect of bexarotene on amyloid- β and apolipoprotein E metabolism in healthy subjects. *Alzheimers Dement. (NY)* 2016, 2, 110-120. PMid:29067298 [View Article](#) [PubMed/NCBI](#)
- [117] Pierrot, N.; Lhommel, R.; Quenon, L.; Hanseeuw, B.; Dricot, L.; Sindic, C.; Maloteaux, J.M.; Octave, J.N.; Ivanoiu, A. Targretin [bexarotene] improves cognitive and biological markers in a patient with Alzheimer's disease. *J. Alzheimer's Dis.* 2016, 49, 271-276. PMid:26444777 [View Article](#) [PubMed/NCBI](#)
- [118] Mirza, Z.; Beg, M.A. Possible molecular interactions of bexarotene - a retinoid drug and Alzheimer's A β peptide: A docking study. *Curr. Alzheimer Res.* 2017, 14, 327-334.
- [119] Huy, P.D.Q.; Thai, N.Q.; Bednarikova, Z.; Phuc, L.H.; Linh, H.Q.; Gazova, Z.; Li, M.S. Bexarotene does not clear amyloid beta plaques but delays fibril growth: Molecular mechanisms. *ACS Chem. Neurosci.* 2017, 8, 1960-1969. PMid:28689412 [View Article](#) [PubMed/NCBI](#)
- [120] Mariani, M.M.; Malm, T.; Lamb, R.; Jay, T.R.; Neilson, L.; Casali, B.; Medarametla, L.; Landreth, G.E. Neuronally-directed effects of RXR activation in a mouse model of Alzheimer's disease. *Sci. Rep.* 2017, 7, 42270. PMid:28205585 [View Article](#) [PubMed/NCBI](#)
- [121] Habchi, J.; Arosio, P.; Perni, M.; Costa, A.R.; Yagi-Utsumi, M.; Joshi, P.; Chia,S.; Cohen, S.I.A.; Muller, M.B.D.; Linse, S.; et al. An anticancer drug suppresses the primary nucleation reaction that initiates the production of the toxic A β 42 aggregates linked with Alzheimer's disease. *Sci. Adv.* 2016, 2, e1501244. PMid:26933687 [View Article](#)

cle [PubMed/NCBI](#)

- [122] Fantini, J.; Di Scala, C.; Yahi, N.; Troadec, J.D.; Sadelli, K.; Chahinian, H.; Garmy, N. Bexarotene blocks calcium-permeable ion channels formed by neurotoxic Alzheimer's β -amyloid peptides. *ACS Chem. Neurosci.* 2014, 5, 216-224. PMid:24383913
[View Article](#) [PubMed/NCBI](#)
- [123] Casali, B.T.; Reed-Geaghan, E.G.; Landreth, G.E. Nuclear receptor agonist-driven modification of inflammation and amyloid pathology enhances and sustains cognitive improvements in a mouse model of Alzheimer's disease. *J. Neuroinflamm.* 2018, 15, 43. PMid:29448961 [View Article](#) [PubMed/NCBI](#)
- [124] Tu, L.; Yang, X.L.; Zhang, Q.; Wang, Q.; Tian, T.; Liu, D.; Qu, X.; Tian, J.Y. Bexarotene attenuates early brain injury via inhibiting microglia activation through PPAR γ after experimental subarachnoid hemorrhage. *Neurol. Res.* 2018, doi:10.1080/01616412.2018.1463900 . PMid:29688151 [View Article](#) [PubMed/NCBI](#)
- [125] Dheer, Y.; Chitranshi, N.; Gupta, V.; Abbasi, M.; Mirzaei, M.; You, Y.; Chung, R.; Graham, S.L.; Gupta, V. Bexarotene modulates retinoid-X-receptor expression and is protective against neurotoxic endoplasmic reticulum stress response and apoptotic pathway activation. *Mol. Neurobiol.* 2018, doi:10.1007/s12035-018-1041-9 . PMid:29637440 [View Article](#) [PubMed/NCBI](#)
- [126] Kamp, F.; Scheidt, H.A.; Winkler, E.; Bassett, G.; Heinel, H.; Hutchison, J.M.; LaPointe, L.M.; Sanders, C.R.; Steiner, H.; Huster, D. Bexarotene binds to the amyloid precursor protein transmembrane domain, alters its α -helical conformation, and inhibits γ -secretase nonselectivity in liposomes. *ACS Chem. Neurosci.* 2018, doi:10.1021acschemneuro.8b00068 .
- [127] Serra-Batiste, M.; Tolchard, J.; Giusti, F.; Zoonens, M.; Carulla, N. Stabilization of a membrane-

associated amyloid- β oligomer for its validation in Alzheimer's disease. *Front. Mol. Biosci.* 2018, 5, 38. PMid:29725595 [View Article](#) [PubMed/NCBI](#)

- [128] Xiang, N.; Lyu, Y.; Zhu, X.; Narsimhan, G. Investigation of the interaction of amyloid- β peptide (11-42) oligomers with a 1-palmitoyl-2-oleoyl-sn-glycero-3-phosphocholine (POPC) membrane using molecular dynamics simulation. *Phys. Chem. Chem.Phys.* 2018, 20, 6817-6829. PMid:29299557 [View Article](#) [PubMed/NCBI](#)
- [129] Habchi, J.; Chia, S.; Galvagnion, C.; Michaels, T.C.T.; Bellaiche, M.M.J.; Ruggeri, F.S.; Sanguani, M.; Idini, I.; Kumita, J.R.; Sparr, E.; et al. Cholesterol catalyses A β 42 aggregation through a heterogeneous nucleation pathway in the presence of lipid membranes. *Nat. Chem.* 2018, 10, 673-683. PMid:29736006 [View Article](#) [PubMed/NCBI](#)
- [130] Bowman, G.L.; Dayon, L.; Kirkland, R.; Wojcik, J.; Peyratout, G.; Severin, I.C.; Henry, H.; Oikonomidi, A.; Migliavacca, E.; Bacher, M.; Popp, J. Blood-brain barrier breakdown, neuroinflammation, and cognitive decline in older adults. *Alzheimer Dementia (online)* 2018, doi:10.1016/j.jalz.2018.06.2857 . PMid:30120040 [View Article](#) [PubMed/NCBI](#)
- [131] Wang, H.; Golob, E.J.; Su, M.Y. Vascular volume and blood-brain barrier permeability measured by dynamic contrast enhanced MRI in hippocampus and cerebellum of patients with MCI and normal controls. *J. Magn. Reson. Imaging* 2006, 24, PMid:16878309 [View Article](#) [PubMed/NCBI](#)
- [132] Montagne, A.; Barnes, S.R.; Sweeney, M.D.; Halliday, M.R.; Sagare, A.P.; Zhao, Z.; Toga, A.W.; Jacobs, R.E.; Liu, C.Y.; Amezcuia, L.; et al. Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* 2015, 85, 296-302. PMid:25611508 [View Article](#) [PubMed/NCBI](#)