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Curcumin and pipernonaline to curb diabetes the natural way: A molecular modeling, docking and dynamics simulation study

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Research

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dium, provided the original work is properly cited.

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ABSTRACT

This study explores natural compounds derived from Indian spices for their antidiabetic activity in search of safe antidiabetics without side effects. Molecular modeling, docking studies followed by replica exchange molecular dynamics simulation studies have been performed on curcumin, trigonelline, pipernonaline and 6-gingerol to study their PPAR γ agonistic activity and α glucosidase inhibition capability, two prominent targets related to manifestation of diabetes mellitus. It is shown that curcumin and pipernonaline form good, stable complexes with the two targets and hence may be useful as nutraceuticals if adopted alongwith diet and other antidiabetic drugs. They may act as preventive measures against diabetes and are totally safe to consume. They may as well be taken as lead compounds for the further development of better, safe antidiabetics.

Keywords: Curcumin; pipernonaline; natural antidiabetics; PPAR gamma; alpha-glucosidase; Safe antidiabetics; molecular docking; modeling; replica exchange molecular dynamics simulations

1. INTRODUCTION

Diabetes: A metabolic disorder

Diabetes mellitus (DM) is one of the most prevalent metabolic disorders around the world. Type 1DM is an autoimmune disease when our body's immune system destroys the beta cells of pancreas resulting in very little insulin production and hence high blood sugar levels. Type 2 DM which accounts for 80% prevalent cases, occurs when our body fails to respond to normal insulin levels, that is becomes insulin resistant (Berg et al, 2002; Moller, 2001) and hence again results in high blood sugar levels. The long term effects are fatal including myocardial infarction, vascular diseases, stroke etc (www.betterhealth.vic.gov.au). A variety of synthetic drugs are available largely based on targets involved in glucose homeostasis. Clinically used drugs include metformin, sulfonyl ureas, α -glucosidase inhibitors, modulators of sodium glucose cotransporter, thiazolidinediones as PPARy agonists, GLP-1R agonists and dipeptidyl peptidase 4 inhibitors etc. (Stein et al, 2013). However these synthetic drugs are associated with some side effects (Lee et al, 2019). Rapid increase in number of T2DM patients motivates us to search natural, safe, non-toxic remedies devoid of side effects. Many plant derived natural compounds with antidiabetic property are being used worldwide (Trojan-Rodrigues et al, 2011; Coman et al, 2012). However, natural compound based therapy has not come up as a standalone treatment option. The lack of knowledge about their mode of action and the fact that many natural compounds are multi targeted are some of the barriers in their effective utilization.

In this study we have considered some natural compounds derived from common Indian spices known to possess some antidiabetic activity and tried to evaluate their potential as antidiabetics. It is with the aim that this work may lead to a supplemental, nontoxic, natural antidiabetic to go along with normal diet; we have taken main constituents of fenugreek, black pepper, ginger and turmeric and attempt to understand their Peroxisome proliferator activated receptor (PPAR γ) agonistic activity and α -glucosidase inhibition capability. PPAR γ is one of the important targets in metabolic disorders to be explored especially if the drug is expected to be multi-targeted. PPAR γ regulates adipogenesis and glucose uptake (Zhou et al, 2015). Many flavonoids and similar compounds derived from natural sources are known to target PPAR γ (Kuroda et al, 2005) and α glucosidase (Muraoka et al, 2010; Ieyama et al, 2011; Feng et al, 2011; Chan et al, 2010). However, their mode of action has not been elucidated at the molecular level. An atomistic level understanding would definitely help design a non-toxic, antidiabetic compound from natural leads.

2. METHODOLOGY

The main constituents of some Indian spices (Niranjan and Prakash, 2008; Grzanna et al, 2005) considered in this study are shown in fig.1. The geometry of these compounds was completely optimized utilizing ab initio quantum mechanical calculations at the Hartree-Fock level with 6-31G basis set (Ditchfield et al, 1971). The optimized conformations are also shown in fig 1. These lowest energy conformations may or may not be the bioactive conformations depending on the conformational flexibility of the compound. The X-ray coordinates of two important targets involved in glucose homeostasis namely PPARy (pdb file: 4A4V) (De Groot et al, 2013) and α -glucosidase (pdbfile: 5NN8) (Roig-Zamboni et al, 2017) were taken from RCSB protein data bank. The proteins were prepared by adding polar hydrogens using Auto Dock version 4 (Morris et al, 1998) and Auto Dock Vina Version 1.1.2 (Trott and Olson, 2010). Grid for docking ligands was generated and authenticated by redocking the ligand present in X-ray crystal structure with an overall RMSD less than 4Å. Ligands prepared for study were then docked one by one in both the active sites (i.e. PPAR γ and α -glucosidase active sites) and their interactions with the active site residues were analysed in detail. AutoDock Vina was used to perform flexible ligand docking studies. Best poses thus obtained were then exported to Shaw's Desmond software (Bowers et al, 2006) to perform replica exchange simulation studies for better and accelerated sampling of conformational space (Earl and Deem, 2005). Replica exchange molecular dynamics (REMD) simulation studies were performed on best

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binding energy complexes to judge the dynamical behaviour and stability of complex with time. All atom SPC water was incorporated in a grid of 10x10x10 Å around the protein and orthorhombic boundary conditions were applied. The minimized complex was subjected to simulated annealing followed by dynamics at 300K for 50ns. Parallel trempering swaps between three adjacent replicas were allowed. NPT ensemble was chosen utilizing Martyna-Tobias-Klein barostat method (Martyna et al, 1994). The RESPA integrator was used with 2fs time step for near and bonded and 6fs time step for far atoms (Tuckerman et al, 1992). Coulombic interaction cutoff was set at 9 Å. RMSD analysis has been performed for all heavy atoms together to judge the stability of the system overtime and fluctuations in position of ligand. Deviations have been taken with respect to initial frame for dynamics.

3. RESULTS AND DISCUSSION

The structures and respective optimized conformations of selected natural compounds are shown in fig. 1. These are largely hydrophobic compounds with some H-bond donor or acceptor groups except trigonelline which occurs in zwitterionic form. After preparing the ligands they were first docked in PPARy active site. The grid placement in active site is shown in fig. 2. First of all the ligand amorfrutin 2 present in X-ray was redocked to authenticate the grid placement. The best pose interactions of redocked amorfrutin are shown in fig. 3. Docking results for top poses are given in table 1. The ionized carboxylic group of amorfrutin can be seen interacting with arg 288 and cys 285. The hydroxyl group of amorfrutin is also H-bonded to neighbouring polar residues. The hydrophobic chain of amorfrutin interacts with neighbouring hydrophobic residues: gly 284, ile 281, ile341 etc. Fig. 4 shows best poses for interactions of curcumin and 6-gingerol in PPARy active site. The methoxy groups on curcumin electrostatically interact with arg 288. The -OH groups are also H-bonded to neighbouring hydrophobic residues lining the active site. Detailed contact analysis is presented in table 2.



Fig. 1 Chemical structures and optimized conformations of natural antidiabetic compounds



Fig. 2 Grid box placement at the active site of PPAR gamma



Fig. 3 Closeup view of redocked pose of amorfrutin 2 in PPAR gamma showing interactions with neighbouring residues

Redocked amorfrutin 2 Curcum					n		6-gingerol			Pipernonaline			Trigonelli	ine	
		Distance from best pose (RMSD in Å)		Distar best p (RMS		from e in Å)		Distance from best pose (RMSD in Å)			Distance from best pose (RMSD in Å)			Distance from best pose (RMSD in Å)	
Pose	Affinity	RMSD Lower bound	RMSD Upper bound	Affinity	RMSD Lower bound	RMSD Upper bound	Affinity	RMSD Lower bound	RMSD Upper bound	Affinity	RMSD Lower bound	RMSD Upper bound	Affinity	RMSD Lower bound	RMSD Upper bound
1	-7.4	0.000	0.000	-8.3	0.000	0.000	-7.3	0.000	0.000	-8.2	0.000	0.000	-4.8	0.000	0.000
2	-7.3	3.427	8.918	-8.1	2.207	2.938	-7.3	1.663	2.714	-7.6	2.479	9.009	-4.8	1.245	2.181
3	-7.2	2.540	4.320	-8.0	7.735	10.435	-7.3	1.051	2.016	-7.4	3.021	9.448	-4.8	3.219	4.143
4	-6.9	2.786	4.962	-7.8	7.961	11.595	-7.1	1.877	2.731	-7.1	24.125	26.029	-4.6	3.099	4.193
5	-6.9	3.536	5.410	-7.7	7.853	10.724	-6.3	2.437	3.874	-7.0	18.086	22.141	-4.6	4.918	5.666
6	-6.8	2.308	6.555	-7.4	2.973	3.718	-6.3	4.571	7.239	-6.7	2.338	8.049	-4.6	4.532	5.193
7	-6.8	3.256	5.779	-7.4	7.957	11.750	-6.3	3.510	7.514	-6.7	18.789	21.908	-4.5	5.862	6.785
8	-6.7	2.917	8.111	-7.3	7.576	11.412	-6.3	3.674	7.301	-6.6	24.779	26.804	-4.4	5.592	6.919
9	-6.7	3.507	5.509	-7.0	6.963	11.046	-6.2	1.739	2.426	-6.6	2.462	8.869	-4.4	14.200	15.137

 Table 1. Docking results of natural antidiabetic compounds in PPAR gamma



Fig. 4 Interactions of curcumin (left) and 6-gingerol (right) with PPAR gamma active site residues

In 6-gingerol the hydrophobic chain of ligand engages the neighbouring hydrophobic residues of active site which are listed in table 2. The –OH at 6-position interacts with arg 288 though not as efficiently as in curcumin. The substituents on the aromatic ring are H -bonded with the backbone of neighbouring ser 342. Binding affinity observed is comparable to amorfrutin (table1) implying the possibility of activating PPAR γ . Docking results for pipernonaline and trigonelline are shown in fig. 5. Trigonelline is held in active site through strong electrostatic interactions with arg 288. Other than electrostatic interactions it is not able to make efficient hydrophobic interactions being very small in size. Its binding affinity (table 1) is also significantly lower than other ligands considered. It is therefore not suited for targeting PPARy. In pipernonaline the ring oxygens interact electrostatically with arg 288. The hydrophobic chain and the alicyclic ring at the other end interact effectively with the largely hydrophobic pocket of the active site. This is reflected in the binding affinity observed (table 1) comparable to amorfrutin and curcumin. Therefore, out of all the natural anti-diabetics considered curcumin and pipernonaline seem to be the best leads to target PPARy. It is understood that interactions must be proper for agonism. Considering the similarity of interactions of curcumin/ pipernonaline with PPARy to amorfrutin with PPARy; curcumin and pipernonaline are suggested as good, natural, non-toxic lead compounds for the development of safe antidiabetics.

Table 2. Detailed contact analysis for best pose of different ligands						
Ligand	Interacting residues in PPAR gamma active site					
AMORFRUTIN 2	ILE281, GLY 284, CYS 285, ARG288, LEU330, ILE341, MET348					
CURCUMIN	ILE262, ARG288, ILE326, MET329, LEU330, LEU333, ILE341					
PIPERNONALINE	ILE281, CYS285, ARG288, ALA292, VAL339, ILE341, SER342					
6-GINGEROL	ARG288, SER289, ALA292, LEU330, ILE326, SER342					
TRIGONELLINE	ARG288, ILE326					



Fig. 5 Interactions of pipernonaline (left) and trigonelline (right) with PPAR gamma active site residues

Apart from PPARy agonistic activity this study also investigates the α -glucosidase inhibition capability of these natural compounds. As per procedure mentioned in methodology section, the active site of α glucosidase was prepared. The drug acarbose present in X-ray structure was redocked to authenticate the grid placement. The best pose interactions of redocked acarbose are presented in fig. 6. The results for top poses are given in table 3. The multiple hydroxyl groups of acarbose can be seen H-bonded with neighbouring asp groups of catalytic domain. Complete list of close contacts is given in table 4. In conformity with previous studies (Monique et al, 1991) stating the importance of asp 518 and trp 516; these residues can be seen H-bonded to acarbose. Fig. 7 shows interactions of curcumin and 6-gingerol with active site residues of a-glucosidase. Curcumin interacts similar to acarbose in active site and hence shows

similar binding affinity. Instead of asp 518, it utilizes asp 616, trp 481 for anchoring to the active site. The two aromatic rings of curcumin form hydrophobic interactions with neighbouring phenylalanine residues. The binding affinity and interactions indicate the α -glucosidase targeting ability of curcumin. The conformation of 6-gingerol is not appropriate for the active site of α -glucosidase which is evident from its lower binding affinity as well (table 3). 6-gingerol is anchored mainly via H-bonding with trp 613, trp 516 and trp 376. It may target α -glucosidase but not as effectively as curcumin. The docking results for pipernonaline and trigonelline are shown in fig. 8. The benzodioxol ring oxygen of pipernonaline is Hbonded to asp 518. The benzodioxol ring also shows pi-pi stacking with phe 649 and trp 481 aromatic rings. The linker connecting the two aromatic rings in pipernonaline undergoes hydrophobic interactions with neighbouring hydrophobic residues lining the active site. Pipernonaline interacts effectively with aglucosidase active site residues and shows binding affinity comparable to curcumin and acarbose. It is therefore suited to target α -glucosidase. Trigonelline is a small molecule and does not occupy entire active site but can be seen anchored utilizing H-bonding with trp 516, trp 481 and arg 600. It is encouraging to see that it also blocks the catalytically important trp 481 residue. It implies that it can certainly inhibit α glucosidase although it may not be highly potent. Again for a-glucosidase inhibition curcumin and pipernonaline seem to be the best natural lead compounds.

Table 3. Docking results of natural antidiabetic compounds in alpha glucosidase															
Redocked Acarbose Curcumin						6-gingerol			Pipernonaline			Trigonelline			
		Distan best (RMS	ce from pose D in Å)	Distance from best pose (RMSD in Å)			Distance from best pose (RMSD in Å)			Distance from best pose (RMSD in Å)		Distance from best pose (RMSD in Å		ce from pose D in Å)	
Pose	Affinity	RMSD Lower bound	RMSD Upper bound	Affinity	RMS D Lower bound	RMSD Upper bound	Affinity	RMS D Lower bound	RMS D Upper bound	Affinity	RMS D Lower bound	RMS D Upper bound	Affinity	RMS D Lower bound	RMS D Upper bound
1	-8.0	0.000	0.000	-7.5	0.000	0.000	-5.6	0.000	0.000	-6.7	0.000	0.000	-4.9	0.000	0.000
2	-7.6	3.199	5.736	-7.4	4.865	5.646	-5.5	1.686	2.069	-6.7	4.099	6.027	-4.9	2.653	3.767
3	-6.8	3.251	5.206	-7.4	3.961	10.392	-5.5	2.514	3.392	-6.6	1.316	2.014	-4.6	1.017	1.489
4	-6.5	3.714	6.076	-7.3	4.089	10.087	-5.4	2.537	6.190	-6.6	5.732	7.936	-4.6	2.873	4.116
5	-6.2	3.682	10.806	-6.7	2.615	2.812	-5.3	2.906	7.331	-6.5	2.501	8.014	-4.3	2.764	4.413
6	-6.1	2.977	11.220	-6.7	1.480	10.313	-5.3	2.492	3.379	-6.5	3.858	8.974	-4.1	2.653	4.186
7	-6.0	6.128	9.293	-6.5	1.442	1.784	-5.3	2.966	4.665	-6.5	5.005	7.750	-4.0	3.262	4.491
8	-5.9	3.117	10.733	-6.4	1.846	9.869	-5.2	2.626	4.444	-6.4	1.568	1.901	-4.0	2.942	3.946
9	-5.8	3.941	10.445	-6.4	6.666	8.470	-5.2	2.640	4.327	-6.4	6.364	9.556	-3.8	7.640	8.563

Table 4. Detailed contact analysis for best pose of different ligands						
Ligand	Interacting residues in alpha glucosidase active site					
ACARBOSE	ARG281, ASP282, ASP404, TRP481, TRP516, ASP518, PHE 525, ARG600, ASP616					
CURCUMIN	ILE441, TRP481, PHE525, TRP613, ASP616, PHE649					
PIPERNONALINE	ARG411, TRP481, ASP518, PHE649, LEU677, LEU678					
6-GINGEROL	TRP376, TRP516, TRP613, PHE649, LEU650					
TRIGONELLINE	TRP481, TRP516, ARG600					



Fig. 6 Closeup view of redocked pose of acarbose in alpha glucosidase showing interactions with neighbouring residues



Fig. 7 Interactions of curcumin (left) and 6-gingerol (right) with alpha glucosidase active site residues

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Fig. 8 Interactions of pipernonaline (left) and trigonelline (right) with alpha glucosidase active site residues

We have also investigated the stabilities of best pose complexes of curcumin and pipernonaline in PPARy and α -glucosidase through REMD simulation studies. Standard MD protocol was followed as mentioned in methodology with three neighbouring replicas. These results are shown in figs. 9 to 11. 50ns trajectories were run in each case. Fig. 9 shows temperature distribution of replicas in temperature spaces for curcumin in PPARy active site. Fig. 10 indicates good stability of both curcumin and pipernonaline in PPARy active site. Fig. 11 shows similar results in aglucosidase. The overall RMSD for all heavy atoms is well below 3Å indicating low fluctuations in position of ligands and good stability of complexes over time for each replica random walk in temperature spaces.



Fig. 9 Temperature distribution for each replica for curcumin in PPAR gamma active site

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Fig. 10 Root mean square deviation for all heavy atoms with respect to initial frame for best pose complex of curcumin (left) and pipernonaline (right) in PPAR gamma active site



Fig. 11 Root mean square deviation for all heavy atoms with respect to initial frame for best pose complex of curcumin (left) and pipernonaline (right) in alpha glucosidase active site

In this study we have shown at the molecular level how curcumin and pipernonaline can target PPAR γ and α -glucosidase similar to other flavonoids and exert antidiabetic action. It is therefore, highly recommended to use curcumin and pipernonaline as nutraceuticals regularly alongwith diet and other antidiabetic drugs to prevent diabetes from worsening and as a preventive measure against its many adverse effects on health.

CONCLUSIONS

This study shows at the molecular level the antidiabetic action of two natural, safe compounds curcumin and pipernonaline derived from Indian spices. It is shown that both form stable complexes with PPAR γ and α -glucosidase active sites, the two important targets related to diabetes. It is therefore recommended to adopt a nutraceutical made out of curcumin and pipernonaline as a dietary supplement to prevent and overcome the adverse effects of diabetes mellitus type 2. It is also recommended to develop a natural, safe antidiabetic drug from these natural, safe lead compounds.

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